

Lecture 2: DNA Microarray Overview

(Some slides from Dr. Holly Dressman, Duke University
http://genome.genetics.duke.edu/STAT_talk_301.ppt)

Announcements

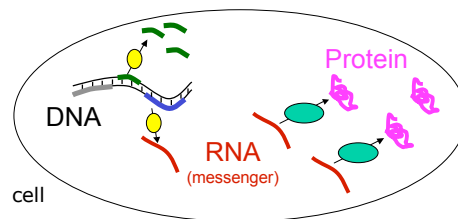
- Go to class web page
<http://www.cs.washington.edu/527>
 - Add yourself to class list
 - Check out HW1, including last year's
- CSE 590CB Org. meeting today,
3:30 MEB 243
<http://www.cs.washington.edu/590cb>

Talks

- **Dr. Martin Tompa**, UW "Tools for Prediction of Regulatory Elements in Microbial Genes"
 - Combi Seminar: Wed 10/6 1:30, K-069
 - CSE Seminar: Tue 10/12 3:30, EE-105
- **Dr. Michal Linial**, Hebrew University, "The Protein Family Tree: Making Biological Sense From Sequence Data"
 - CSE Seminar: Thu 10/7 3:30 pm, EE-105

Gene Expression: The "Central Dogma"

DNA → RNA → Protein

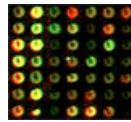


Gene Expression

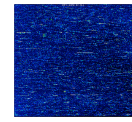
- Proteins do most of the work
- They're dynamically created/destroyed
- So are their mRNA blueprints
- Different mRNAs expressed at different times/places
- Knowing mRNA "expression levels" tells a lot about the state of the cell

Microarrays

A snapshot that captures the activity pattern of thousands of genes at once.



Custom spotted arrays

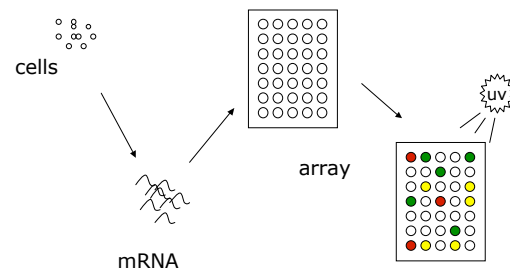


Affymetrix GeneChip

Expression Microarrays

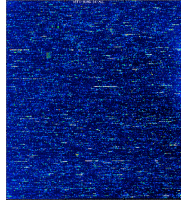
- The Array
 - Thousands to hundreds of thousands of spots per square inch
 - Each holds millions of copies of a DNA sequence from one gene
- Its Use
 - Take mRNA from cells, put it on array
 - See where it sticks – mRNA from gene x should stick to spot x

An Expression Array Experiment



An Example Application

- 72 leukemia patients
 - 47 ALL
 - 25 AML
- 1 chip per patient
- 7132 human genes per chip



Golub, et al., Science 286:531-537 (1999).

Key Issue: What's Different?

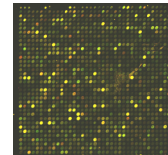
- What genes are behaving differently between ALL & AML (or other disease/normal states)?
- Potential uses:
 - Diagnosis
 - Prognosis
 - Insight into underlying biology/biologies
 - Treatment

A Classification Problem

- Given an array from a new patient: is it ALL or AML?
- Many possible approaches: LDA, logistic regression, NN, SVM, ...
- Problems:
 - Noise
 - Dimensionality

An Example Application

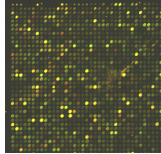
- Yeast "Sporulation"
- 7 time points over ~18 hours
- One array per time point
- All 6200 yeast genes on each



Chu, DeRisi, Eisen, Mulholland, Botstein, Brown, Herskowitz, "The Transcriptional Program of Sporulation in Budding Yeast," Science, 282 (Oct 1998) 699-705

An Example Application

- Yeast “Sporulation”
- 7 time points over ~18 hours
- One array per time point
- All 6200 yeast genes on each



3-10x increase in number of genes known to be involved in sporulation, many with recognizable analogs in humans, presumably key players in egg/sperm formation

Chu, DeRisi, Eisen, Mulholland, Botstein, Brown, Herskowitz, "The Transcriptional Program of Sporulation in Budding Yeast," Science, 282 (Oct 1998) 699-705

Other Applications

- Study gene function & regulation
 - Covarying \rightsquigarrow coregulated?
 - Covarying \rightsquigarrow common pathway?
- Refined categorization of diseases
 - E.g., "prostate cancer" is almost certainly not one disease. Are subtypes distinguishable at expression level?

Practical Applications of Microarrays

Gene Target Discovery

- Diseased vs normal cell comparison suggests sets of genes having key roles.
- Over/underexpressed genes in the diseased cells can suggest drug targets

Pharmacology and Toxicology

- Highly sensitive indicator of a drug's activity (pharmacology) and toxicity (toxicology) in cell culture or test animals.
- Screen or optimize drug candidates prior to costly clinical trials.

Diagnostics

- Potential to diagnose clinical conditions by detecting gene expression patterns associated with disease states in either biopsy samples or peripheral blood cells.

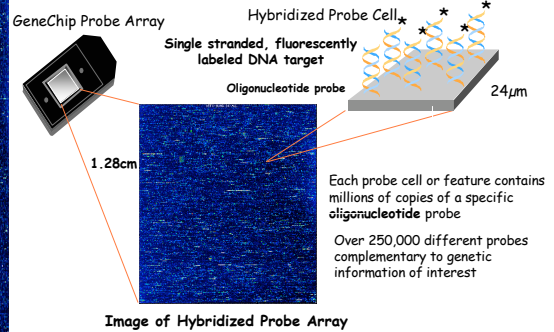
Microarray Technologies

- Oligo Arrays
 - Affymetrix -
 - one color
 - Short oligos
 - match/mismatch
 - Agilent, inter alia
 - 2 color
 - Longer oligos
- Spotted cDNA arrays

GeneChip® Probe Array



GeneChip® Probe Arrays



How unique is a 20-mer?

- VERY CRUDE model: DNA is random—every position is equally likely to be A, C, G, or T, independent of every other

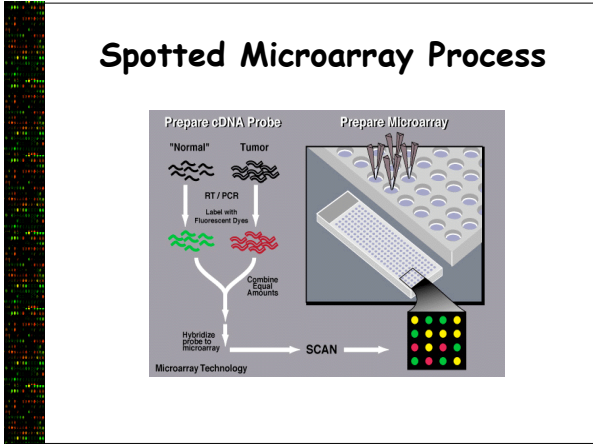
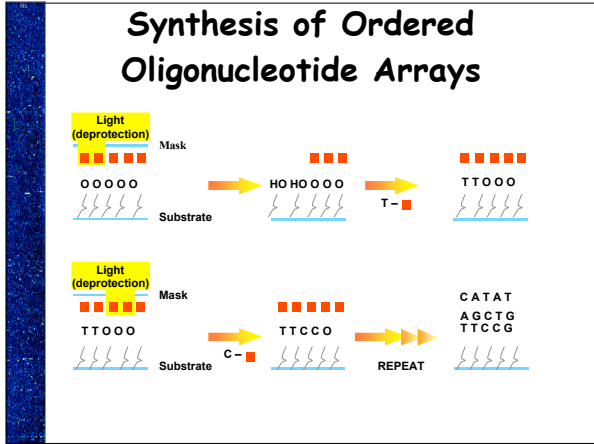
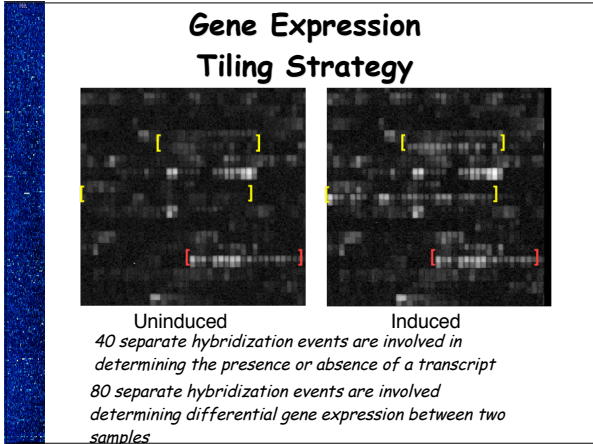
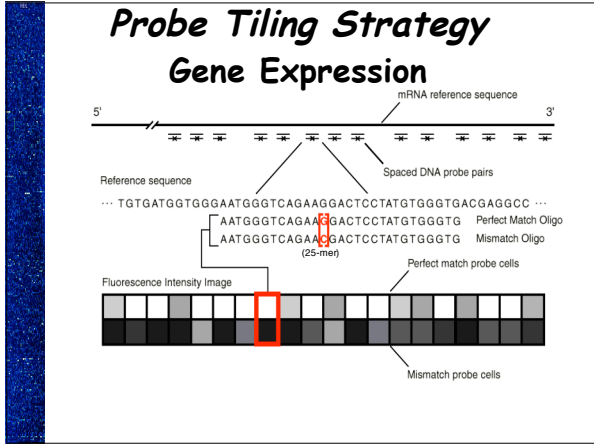
- Then probability of a random 20-mer is

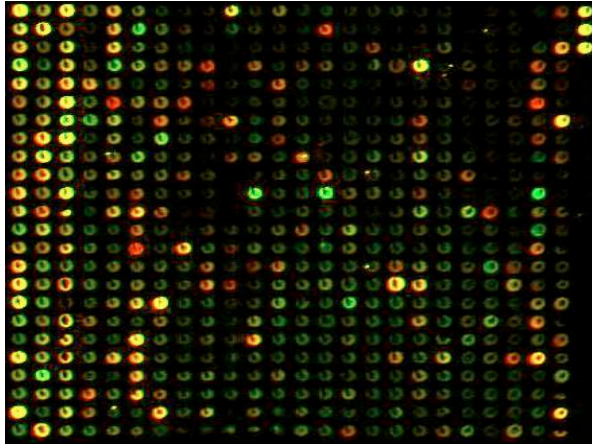
$$\left(\frac{1}{4}\right)^{20} = \left(\frac{1}{2}\right)^{40} = \left(\left(\frac{1}{2}\right)^{10}\right)^4 = \left(\left(\frac{1}{1024}\right)\right)^4 \approx (10^{-3})^4 = 10^{-12}$$

- So, a specific 20-mer occurs in random human-sized DNA sequence with probability about $3 \times 10^9 \times 10^{-12} = .003$

How Random is a Genome?

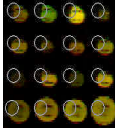
- G/C content can vary from ~40-60% across and within organisms ("isochores")
- Adjacent pairs not independent
- Adjacent triples not independent (esp. in genes)
- ...
- Many large-scale repeats, e.g.
 - similar genes, domains within genes
 - transposons & other junk
 - within primates, ~5% of all DNA is composed of (noisy) copies of a 300bp ALU sequence
- Nevertheless, crude model above is a useful guide



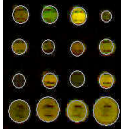


GenePix Pro Features




- Auto Align



Before Auto Align



After Auto Align

Ratio (635 nm/532 nm)	
	Rp: 1.238 Rm: 1.056 mR: 1.106 rR: 2.023
Wavelength 635 nm	
	P: 13609 F: 12199 B: 135
Wavelength 532 nm	
	P: 10992 F: 11534 B: 105
Pixels: F=80, B=420	

P = pixel intensity
 F = feature intensity
 B = background intensity
 Rp = ratio of pixel intensities
 Rm = ratio of means
 mR = median of ratios
 rR = regression ratio

Spotted glass slide microarrays

Advantages

- Low cost per array
- Custom gene selection
- Any species
- Competitive hybridization
- Open architecture

Disadvantages

- Clone management
- Clone cost
- Quality control

Affymetrix GeneChip system

Advantages

Stream line production
Large number of genes and ESTs/chip
Several number of species

Disadvantages

System cost
GeneChip cost
Proprietary system
Limits on customizing

Micro Array Noise Sources

- Lot-to-lot variation (chips, reagents,...)
- Experiment-to-experiment variation
 - cell state, culture purity
 - sample preparation, hybridization conditions
- Spot-to-spot variation
 - unequal dye incorporation
 - dye nonlinearity/saturation
 - uneven spot sizes
 - self- & cross-hybridization
 - Image capture & processing (spot finding, quantization, sensors)
- ...

Challenges in analyzing Microarray Data

- Amount of DNA in spot is not consistent
- Spot contamination
- cDNA may not be proportional to that in the tissue
- Low hybridization quality
- Measurement errors
- Spliced variants
- Outliers
- Data are high-dimensional "multi-variant"
- Biological signal may be subtle, complex, non linear, and buried in a cloud of noise
- Normalization
- Comparison across multiple arrays, time points, tissues, treatments
- How do you reveal biological relationships among genes?
- How do you distinguish real effect from artifact?

Factors to consider in designing microarray experiments

- Need to do lots of control experiments-validate method
- Do replicate spotting, replicate chips, and reverse labeling for custom spotted chips
- Do pilot studies before doing "mega chip" experiments
- Don't design experiment without replication; nothing will be learned from a single failed experiment
- Design simple (one-two factor) experiments, i.e. treatment vs. untreated
- Understand measurement errors
- In designing Databases; they are useful ONLY if quality of data is assured
- Involve statistical colleagues in the design stages of your studies

Microarray Summary

- Lots of variations
 - Glass, nylon
 - Long, short DNA molecules
 - Fab via photolithography, ink jet, robot
 - Radioactive vs fluorescent readout
 - Relative vs absolute intensity
- Leads to diverse sensitivity, bias, noise, etc.
- But same bottom line:
unprecedented global insight into cellular state and function

The Microarray Biz. (circa 3/2001)

- Despite concerns above...
- "In early 1997, scientists never envisioned looking at more than 25 to 50 gene-expression levels simultaneously. Today everybody tells us that they want to look at the whole genome." -- T.Kreiner, Affymetrics
- 45% annual growth rate 1999-2000