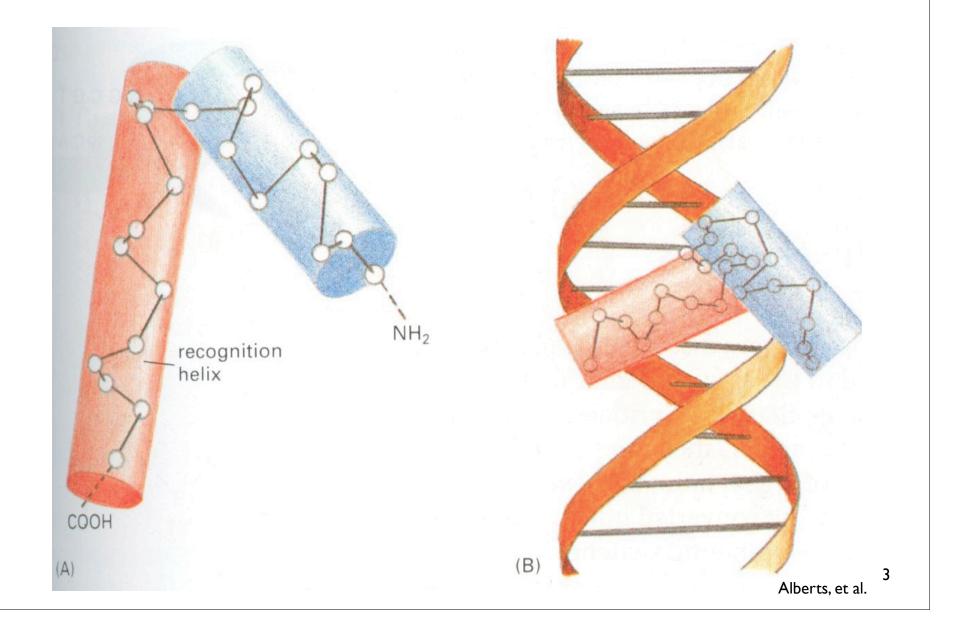
#### CSEP 590A Summer 2006 Lecture 5 Motifs: Representation & Discovery

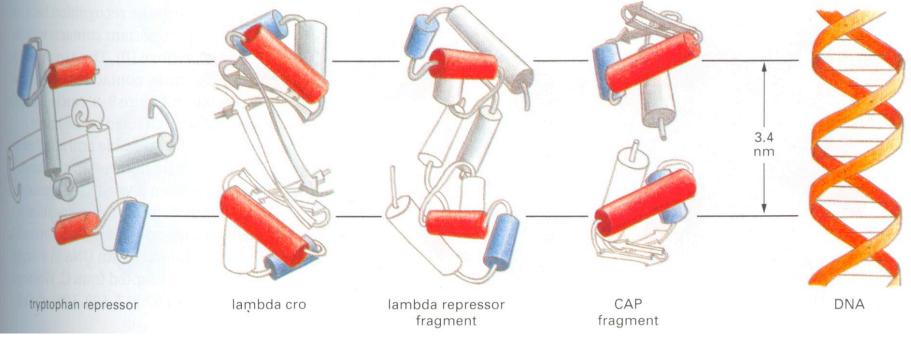
# **DNA Binding Proteins**

A variety of DNA binding proteins ("transcription factors"; a significant fraction, perhaps 5-10%, of all human proteins) modulate transcription of protein coding genes

#### Helix-Turn-Helix DNA Binding Motif

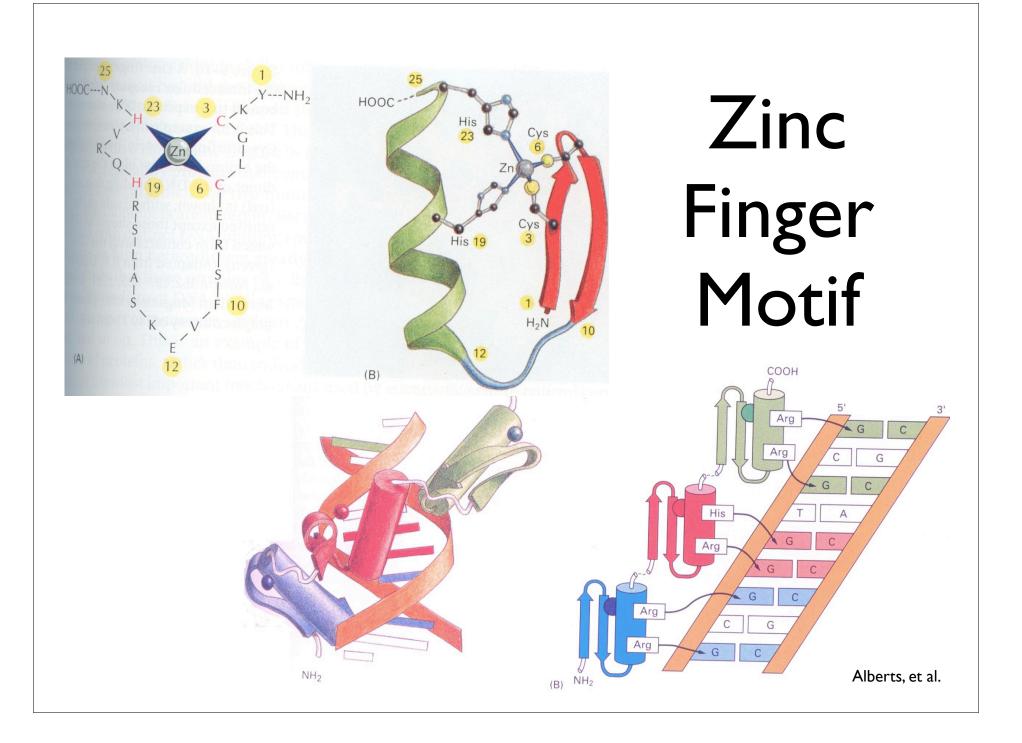


#### H-T-H Dimers

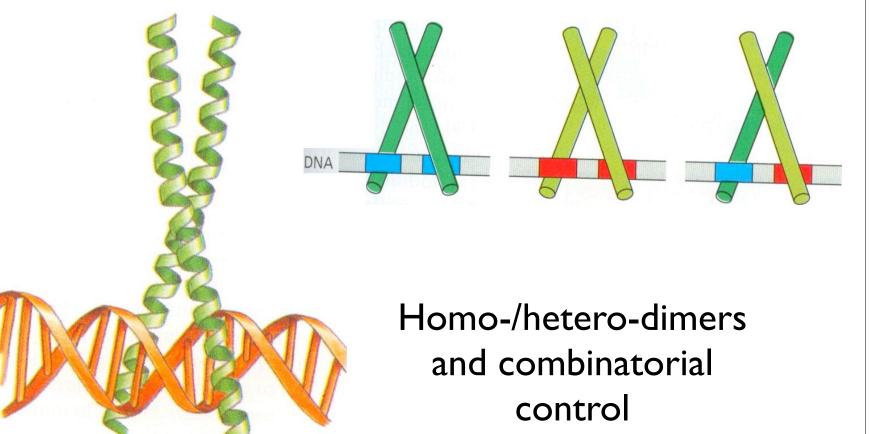


Alberts, et al.

#### Bind 2 DNA patches, ~ I turn apart Increases both specificity and affinity



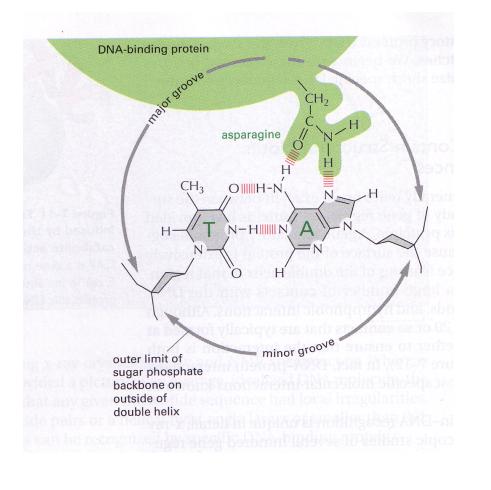
# Leucine Zipper Motif

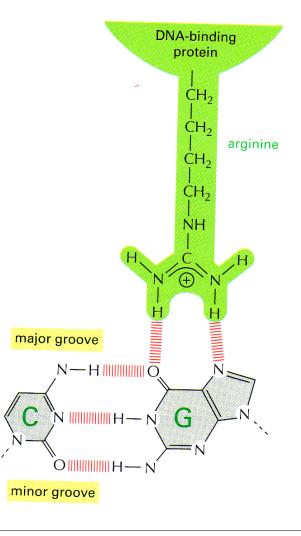


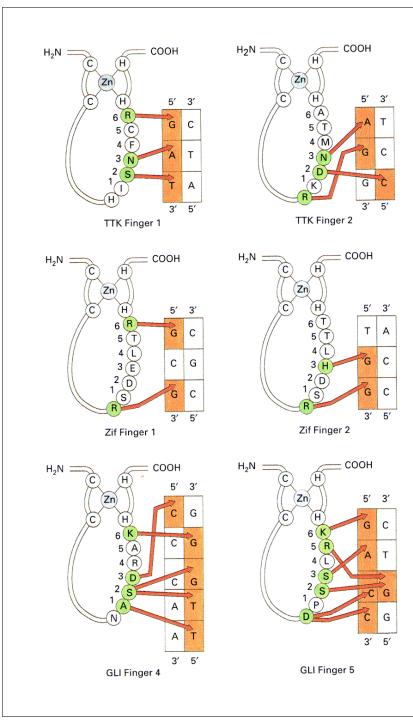
Alberts, et al.

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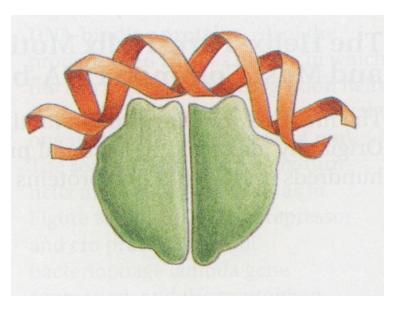
#### Some Protein/DNA interactions well-understood







But the overall DNA binding "code" still defies prediction



# DNA binding site summary

- complex "code"
- short patches (6-8 bp)
- often near each other (I turn = I0 bp)
- often reverse-complements
- not perfect matches

#### Sequence Motifs

#### E. coli Promoters

- "TATA Box" ~ 10bp upstream of transcription start
- How to define it?
  - Consensus is TATAAT
  - BUT all differ from it
  - allow k mismatches?
  - equally weighted?

TACGAT TAAAAT TATACT GATAAT TATGAT TATGTT

• wildcards like R,Y? ({A,G}, {C,T}, resp.)

#### E. coli Promoters

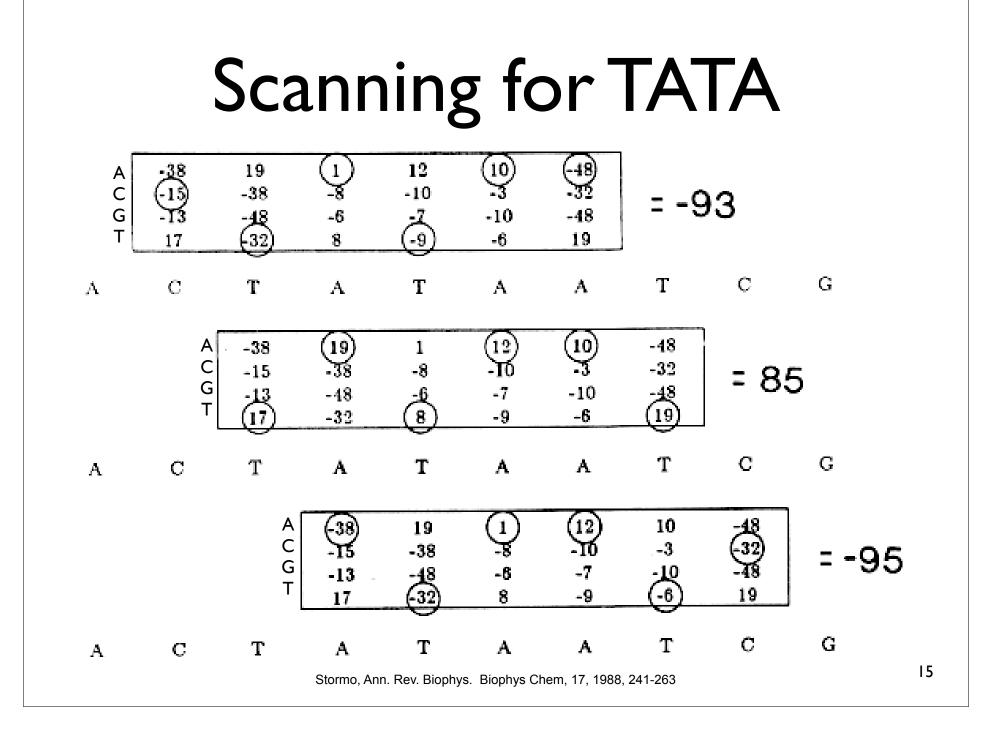
- "TATA Box" consensus TATAAT ~
   10bp upstream of transcription start
- Not exact: of 168 studied (mid 80's)
  - nearly all had 2/3 of TAxyzT
  - 80-90% had all 3
  - 50% agreed in each of x,y,z
  - no perfect match
- Other common features at -35, etc.

#### **TATA Box Frequencies**

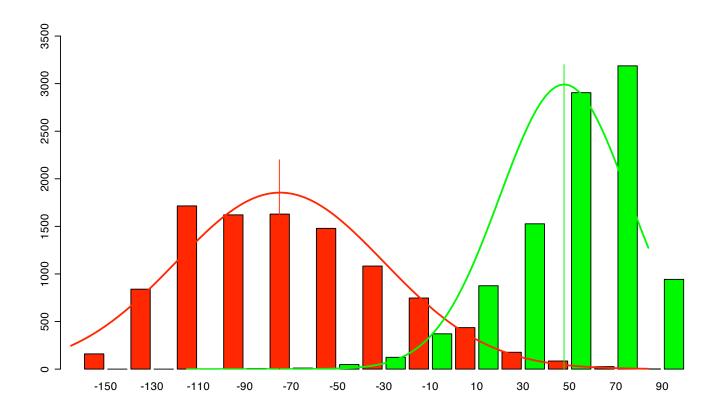
pos base	1	2	3	4	5	6
A	2	95	26	59	51	1
С	9	2	14	13	20	3
G	10	1	16	15	13	0
Т	79	3	44	13	17	96

#### **TATA** Scores

pos base	1	2	3	4	5	6
Α	-36	19	1	12	10	-46
С	-15	-36	-8	-9	-3	-31
G	-13	-46	-6	-7	-9	-46(?)
Τ	17	-31	8	-9	-6	19



#### Score Distribution (Simulated)



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### Weight Matrices: Statistics

• Assume:

 $f_{b,i}$  = frequency of base *b* in position *i* in TATA

 $f_b$  = frequency of base b in all sequences

• Log likelihood ratio, given  $S = B_1 B_2 \dots B_6$ :

$$\log_{-} \left( \frac{P(S|\text{``tata''})}{P(S|\text{``non-tata''})} \right) = \log_{-} \frac{\prod_{i=1}^{6} f_{B_{i},i}}{\prod_{i=1}^{6} f_{B_{i}}} = \sum_{i=1}^{6} \log_{-} \frac{f_{B_{i},i}}{f_{B_{i}}}$$

Assumes independence

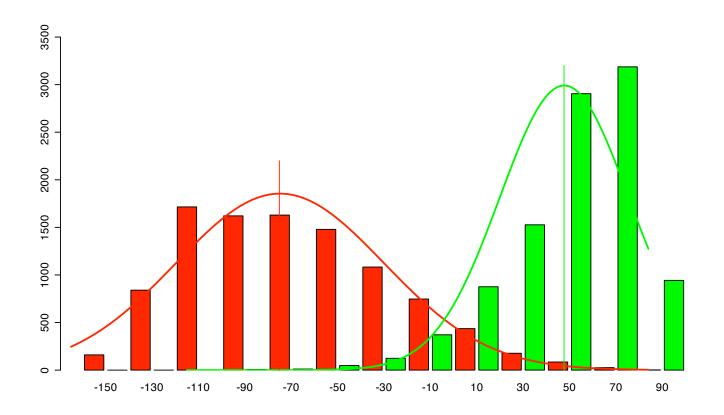
### Neyman-Pearson

- Given a sample  $x_1, x_2, ..., x_n$ , from a distribution  $f(...|\Theta)$  with parameter  $\Theta$ , want to test hypothesis  $\Theta = \theta_1$  vs  $\Theta = \theta_2$ .
- Might as well look at likelihood ratio:

$$\frac{f(x_1, x_2, ..., x_n | \theta_1)}{f(x_1, x_2, ..., x_n | \theta_2)} > \tau$$

(or log likelihood difference)

# Score Distribution (Simulated)



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#### What's best WMM?

- Given 20 sequences s<sub>1</sub>, s<sub>2</sub>, ..., s<sub>k</sub> of length 8, assumed to be generated at random according to a WMM defined by 8 x (4-1) parameters θ, what's the best θ?
- E.g., what's MLE for  $\theta$  given data  $s_1, s_2, ..., s_k$ ?
- Answer: count frequencies per position.

# Weight Matrices: Chemistry

 Experiments show ~80% correlation of log likelihood weight matrix scores to measured binding energy of RNA polymerase to variations on TATAAT consensus [Stormo & Fields]

### Another WMM example

8 Sequences: ATG ATG ATG ATG ATG GTG GTG GTG TTG

Freq.	Col I	Col 2	Col3
Α	.625	0	0
С	0	0	0
G	.250	0	Ι
Т	.125		0

LLR	Col I	Col 2	Col 3
А	I.32	-∞	-∞
С	-∞	-∞	-∞
G	0	-∞	2.00
Т	-1.00	2.00	-∞

Log-Likelihood Ratio:

$$\log_2 \frac{f_{x_i,i}}{f_{x_i}}, \ f_{x_i} = \frac{1}{4}$$

## Non-uniform Background

- E. coli DNA approximately 25% A, C, G, T
- *M. jannaschi* 68% A-T, 32% G-C

LLR from previous example, assuming

$$f_A = f_T = 3/8$$
  
 $f_C = f_G = 1/8$ 

LLR	Col I	Col 2	Col 3
А	.74	-8	-∞
С	-∞	-∞	-∞
G	1.00	-8	3.00
Т	-1.58	I.42	-∞

e.g., G in col 3 is 8 x more likely via WMM than background, so  $(\log_2)$  score = 3 (bits).

#### Relative Entropy

- AKA Kullback-Liebler Distance/Divergence, AKA Information Content
- Given distributions P, Q

$$H(P||Q) = \sum_{x \in \Omega} P(x) \log \frac{P(x)}{Q(x)} \ge \mathbf{0}$$

Notes:

Let 
$$P(x)\log \frac{P(x)}{Q(x)} = 0$$
 if  $P(x) = 0$  [since  $\lim_{y \to 0} y \log y = 0$ ]

Undefined if 0 = Q(x) < P(x)

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#### WMM: How "Informative"? Mean score of site vs bkg?

- For any fixed length sequence x, let
   P(x) = Prob. of x according to WMM
   Q(x) = Prob. of x according to background
- Relative Entropy:

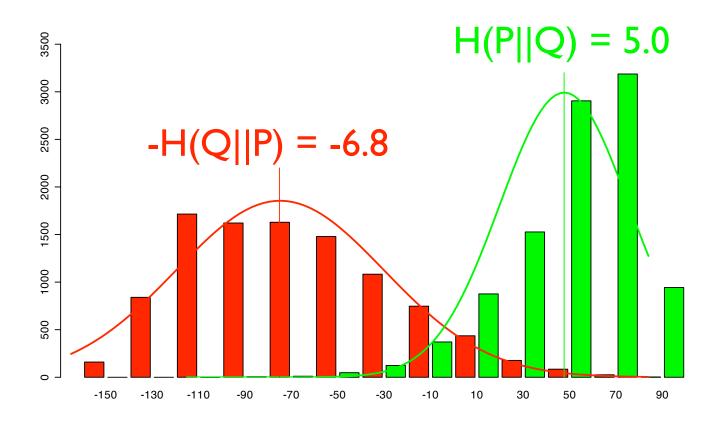
$$H(P||Q) = \sum_{x \in \Omega} P(x) \log_2 \frac{P(x)}{Q(x)}$$

H(P||Q) is expected log likelihood score of a sequence randomly chosen from WMM;
 -H(Q||P) is expected score of Background

-H(Q||P)

H(P||Q)

# WMM Scores vs Relative Entropy



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For WMM, you can show (based on the assumption of independence between columns), that :

 $H(P||Q) = \sum_{i} H(P_i||Q_i)$ 

where  $P_i$  and  $Q_i$  are the WMM/background distributions for column i.

## WMM Example, cont.

Freq.	Col I	Col 2	Col3
Α	.625	0	0
С	0	0	0
G	.250	0	I
T .125			0

#### Uniform

Col I	Col 2	Col 3				
1.32	-8	-8				
-8	- 8	-8				
0	- 8	2.00				
-1.00	2.00	-∞				
.70	2.00	2.00	4.70			
	1.32 -∞ 0 -1.00	$1.32$ $-\infty$ $-\infty$ $-\infty$ $0$ $-\infty$ $-1.00$ $2.00$	$-\infty$ $-\infty$ $-\infty$ 0 $-\infty$ 2.00 -1.00 2.00 $-\infty$			

#### Non-uniform

LLR	Col I	Col 2	Col 3	
Α	.74	-∞	-8	
С	-∞	-∞	-8	
G	1.00	-∞	3.00	
Т	-1.58	1.42	-∞	
RelEnt	.51	1.42	3.00	4.93

#### Pseudocounts

- Are the  $-\infty$ 's a problem?
  - Certain that a given residue never occurs in a given position? Then -∞ just right
  - Else, it may be a small-sample artifact
- Typical fix: add a *pseudocount* to each observed count—small constant (e.g., .5, I)
- Sounds ad hoc; there is a Bayesian justification

# WMM Summary

- Weight Matrix Model (aka Position Specific Scoring Matrix, PSSM, "possum", 0th order Markov models)
- Simple statistical model assuming independence between adjacent positions
- To build: count (+ pseudocount) letter frequency per position, log likelihood ratio to background
- To scan: add LLRs per position, compare to threshold
- Generalizations to higher order models (i.e., letter frequency per position, conditional on neighbor) also possible, with enough training data

### How-to Questions

- Given aligned motif instances, build model?
  - Frequency counts (above, maybe with pseudocounts)
- Given a model, find (probable) instances
  - Scanning, as above
- Given unaligned strings thought to contain a motif, find it? (e.g., upstream regions for co-expressed genes from a microarray experiment)
  - Hard... rest of lecture.

### Motif Discovery

Unfortunately, finding a site of max relative entropy in a set of unaligned sequences is NP-hard [Akutsu]

# Motif Discovery: 3 example approaches

- Greedy search
- Expectation Maximization
- Gibbs sampler

#### Greedy Best-First Approach [Hertz & Stormo]

Input:

Sequence s<sub>1</sub>, s<sub>2</sub>, ..., s<sub>k</sub>; motif length *l*; "breadth" *d* Algorithm:

- create singleton set with each length *l* subsequence of each s<sub>1</sub>, s<sub>2</sub>, ..., s<sub>k</sub>
- for each set, add each possible length *l* subsequence not already present
- compute relative entropy of each
- discard all but d best
- repeat until all have k sequences

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#### Expectation Maximization [MEME, Bailey & Elkan, 1995]

Input (as above):

 Sequence s<sub>1</sub>, s<sub>2</sub>, ..., s<sub>k</sub>; motif length *l*; background model; again assume one instance per sequence (variants possible)

Algorithm: EM

- Visible data: the sequences
- Hidden data: where's the motif

 $Y_{i,j} = \begin{cases} 1 & \text{if motif in sequence } i \text{ begins at position } j \\ 0 & \text{otherwise} \end{cases}$ 

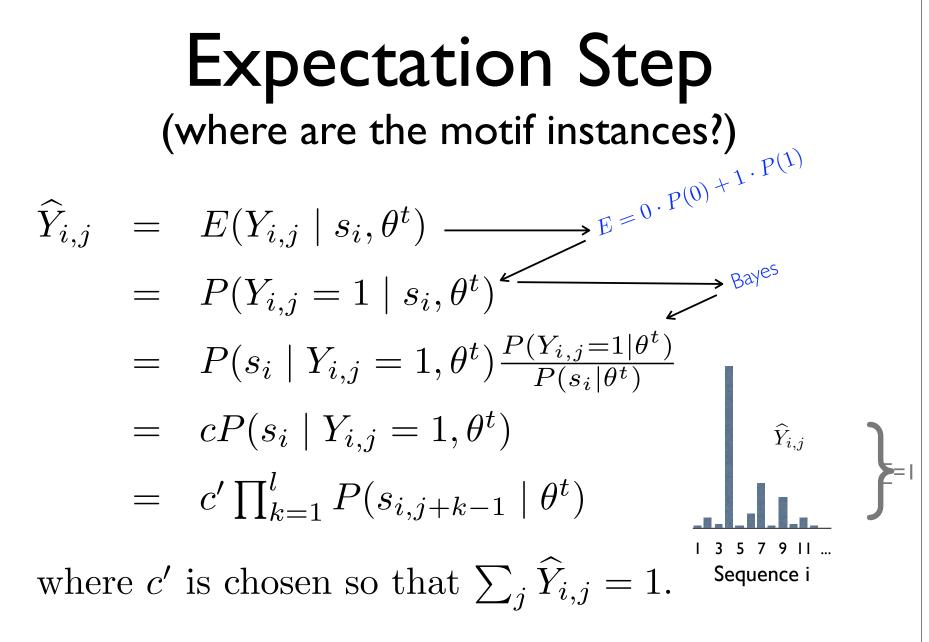
• Parameters  $\theta$ : The WMM

# **MEME** Outline

Typical EM algorithm:

- Parameters  $\theta^t$  at  $t^{th}$  iteration, used to estimate where the motif instances are (the hidden variables)
- Use those estimates to re-estimate the parameters  $\theta$  to maximize likelihood of observed data, giving  $\theta^{t+l}$
- Repeat

Key: given a few good matches to best motif, expect to pick out more



#### Maximization Step (what is the motif?)

#### Find $\theta$ maximizing expected value:

$$\begin{aligned} Q(\theta \mid \theta^{t}) &= E_{Y \sim \theta^{t}} [\log P(s, Y \mid \theta)] \\ &= E_{Y \sim \theta^{t}} [\log \prod_{i=1}^{k} P(s_{i}, Y_{i} \mid \theta)] \\ &= E_{Y \sim \theta^{t}} [\sum_{i=1}^{k} \log P(s_{i}, Y_{i} \mid \theta)] \\ &= E_{Y \sim \theta^{t}} [\sum_{i=1}^{k} \sum_{j=1}^{|s_{i}| - l + 1} Y_{i,j} \log P(s_{i}, Y_{i,j} = 1 \mid \theta)] \\ &= E_{Y \sim \theta^{t}} [\sum_{i=1}^{k} \sum_{j=1}^{|s_{i}| - l + 1} Y_{i,j} \log (P(s_{i} \mid Y_{i,j} = 1, \theta) P(Y_{i,j} = 1 \mid \theta))] \\ &= \sum_{i=1}^{k} \sum_{j=1}^{|s_{i}| - l + 1} E_{Y \sim \theta^{t}} [Y_{i,j}] \log P(s_{i} \mid Y_{i,j} = 1, \theta) + C \\ &= \sum_{i=1}^{k} \sum_{j=1}^{|s_{i}| - l + 1} \widehat{Y}_{i,j} \log P(s_{i} \mid Y_{i,j} = 1, \theta) + C \end{aligned}$$

# M-Step (cont.)

 $Q(\theta \mid \theta^{t}) = \sum_{i=1}^{k} \sum_{j=1}^{|s_{i}|-l+1} \widehat{Y}_{i,j} \log P(s_{i} \mid Y_{i,j} = 1, \theta) + C$ 

Exercise: Show this is maximized by "counting" letter frequencies over all possible motif instances, with counts weighted by  $\hat{Y}_{i,j}$ , again the "obvious" thing.

 $s_1$ : ACGGATT...  $s_k$ : GC...TCGGAC



 $egin{array}{ccc} dots & dots \ \widehat{Y}_{k,l-1} & \mathsf{CGGA} \ \widehat{Y}_{k,l} & \mathsf{GGAC} \end{array}$ 

#### Initialization

- 1. Try every motif-length substring, and use as initial  $\theta$  a WMM with, say 80% of weight on that sequence, rest uniform
- 2. Run a few iterations of each
- 3. Run best few to convergence

(Having a supercomputer helps)

### Another Motif Discovery Approach The Gibbs Sampler

Lawrence, et al. "Detecting Subtle Sequence Signals: A Gibbs Sampling Strategy for Multiple Sequence Alignment," Science 1993

Sigma-37	223	IIDLTYIQNK	SQKETGDILGISQMHVSR	LQRKAVKKLR	240	A25944	
SpoIIIC	94	RFGLDLKKEK	TQREIAKELGISRSYVSR	IEKRALMKMF	111	A28627	
NahR	22	VVFNQLLVDR	RVSITAENLGLTQPAVSN	ALKRLRTSLQ	39	A32837	
Antennapedia	326	FHFNRYLTRR	RRIEIAHALCLTERQIKI	WFQNRRMKWK	343	A23450	
NtrC (Brady.)	449	LTAALAATRG	NQIRAADLLGLNRNTLRK	KIRDLDIQVY	466	B26499	
DicA	22	IRYRRKNLKH	TQRSLAKALKISHVSVSQ	WERGDSEPTG	39	B24328	(BVECDA)
MerD	5	MNAY	TVSRLALDAGVSVHIVRD	YLLRGLLRPV	22	C29010	
Fis	73	LDMVMQYTRG	NQTRAALMMGINRGTLRK	KLKKYGMN	90	A32142	(DNECFS)
MAT al	99	FRRKQSLNSK	EKEEVAKKCGITPLQVRV	WFINKRMRSK	116	A90983	(JEBY1)
Lambda cII	25	SALLNKIAML	GTEKTAEAVGVDKSQISR	WKRDWIPKFS	42	A03579	(QCBP2L)
Crp (CAP)	169	THPDGMQIKI	TRQEIGQIVGCSRETVGR	ILKMLEDQNL	186	A03553	(QRECC)
Lambda Cro	15	ITLKDYAMRF	GQTKTAKDLGVYQSAINK	AIHAGRKIFL	32	A03577	(RCBPL)
P22 Cro	12	YKKDVIDHFG	TQRAVAKALGISDAAVSQ	WKÉVIPEKDA	29	A25867	(RGBP22)
AraC	196	ISDHLADSNF	DIASVAQHVCLSPSRLSH	LFRQQLGISV	213	A03554	(RGECA)
Fnr	196	FSPREFRLTM	TRGDIGNYLGLTVETISR	LLGRFQKSGM	213	A03552	(RGECF)
HtpR	252	ARWLDEDNKS	TLQELADRYGVSAERVRQ	LEKNAMKKLR	269	A00700	(RGECH)
NtrC (K.a.)	444	LTTALRHTQG	HKQEAARLLGWGRNTLTR	KLKELGME	461	A03564	(RGKBCP)
CytR	11	MKAKKQETAA	TMKDVALKAKVSTATVSR	ALMNPDKVSQ	28	A24963	(RPECCT)
DeoR	23	LQELKRSDKL	HLKDAAALLGVSEMTIRR	DLNNHSAPVV	40	A24076	(RPECDO)
GalR	3	MA	TIKDVARLAGVSVATVSR	VINNSPKASE	20	A03559	(RPECG)
LacI	5	MKPV	TLYDVAEYAGVSYQTVSR	VVNQASHVSA	22	A03558	(RPECL)
TetR	26	LLNEVGIEGL	TTRKLAQKLGVEQPTLYW	HVKNKRALLD	43	A03576	(RPECTN)
TrpR	67	IVEELLRGEM	SQRELKNELGAGIATITR	GSNSLKAAPV	84	A03568	(RPECW)
NifA			VQAKAARLLGMTPRQVAY	-	512	S02513	
SpoIIG	205	RFGLVGEEEK	TQKDVADMMGISQSYISR	LEKRIIKRLR	222	S07337	
, Pin	160	QAGRLIAAGT	PRQKVAIIYDVGVSTLYK	TFPAGDK	177	S07958	
PurR	- 3	MA	TIKDVAKRANVSTTTVSH	VINKTRFVAE	20	S08477	
EbgR	3	MA	TLKDIAIEAGVSLATVSR	VLNDDPTLNV	20	S09205	
LexA	27		TRAEIAQRLGFRSPNAAE		44	S11945	
P22 cI	25	SSILNRIAIR	GQRKVADALGINESQISR		42	B25867	(Z1BPC2)
			******	***			

В								Posit	ion i	n site								
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
Arg	94	222	265	137	9	9	137	137	9	9	9	52	222	94	94	9	265	606
Lys	9	133	442	380	9	71	380	194	9	133	9	9	71	9	9	9	71	256
Glu	53	9	96	401	9	9	140	140	9	9	9	53	140	140	9	9	9	53
Asp	67	9	9	473	9	9	299	125	9	67	9	67	67	9	9	9	9	67
Gln	9	600	224	9	9	9	224	9	9	9	9	9	278	63	278	9	9	170
His	240	9	<sup>-</sup> 9	9	9	9	125	125	9	9	9	9	125	125	125	9	9	240
Asn	168	9	9	9	9	9	168	89	9	89	9	248	9	168	89	9	89	89
Ser	117	9	117	117	9	9	9	9	9	9	9	819	63	387	63	9	819	9
Gly	151	9	56	9	9	151	9	9	9	1141	9	151	9	56	9	9	56	9
Ala	.9	9	112	43	181	901	43	181	215	9	43	9	43	181	112	- <u>4</u> 3	78	9
Thr	915	130	130	9	251	9	9	9	9	9	9	311	130	70	855	ີ 9	130	9
Pro	76	9	9	9	9	9	9	9	9	9	9	9	210	210	9	9	9,	9
Cys	9	9	9	9	9	9	9	9	295	581	295	9	9	9	9	9	. 9	9
Val	58	107	9	9	500	9	9	9	156	9	598	9	205	58	9	746	9	58
Leu	9	121	9	9	149	9	93	149	458	9	149	9	37	37	9	177	9	9
Ile	9	166	114	61	323	9	114	166	9	9	427	9.	61	9	61	427	9	61
Met	9	104	9	9	9	9	9	198	198	9	104	9	9	198	9	9	9	9
Tyr	9	9	136	9	· 9	9	9	262	262	9	9	136	136	9	262	9	262	136
Phe	9	9	9	9	9	9	9	9	9	9	108	9	9	9	9	9	9	9
Trp	9	9	9	9	9	9	9	9	9	9	366	9	9	9	9	9	9	366

#### Some History

- Geman & Geman, IEEE PAMI 1984
- Hastings, Biometrika, 1970
- Metropolis, Rosenbluth, Rosenbluth, Teller, & Teller, "Equations of State Calculations by Fast Computing Machines," J. Chem. Phys. 1953
- Josiah Williard Gibbs, 1839-1903, American physicist, a pioneer of thermodynamics

#### How to Average

An old problem:

- n random variables:
- Joint distribution (p.d.f.):
- Some function:
- <u>Want</u> Expected Value:

 $x_1, x_2, \dots, x_k$  $P(x_1, x_2, \dots, x_k)$  $f(x_1, x_2, \dots, x_k)$  $E(f(x_1, x_2, \dots, x_k))$ 

### How to Average

$$E(f(x_1, x_2, \dots, x_k)) = \int_{x_1} \int_{x_2} \dots \int_{x_k} f(x_1, x_2, \dots, x_k) \cdot P(x_1, x_2, \dots, x_k) dx_1 dx_2 \dots dx_k$$

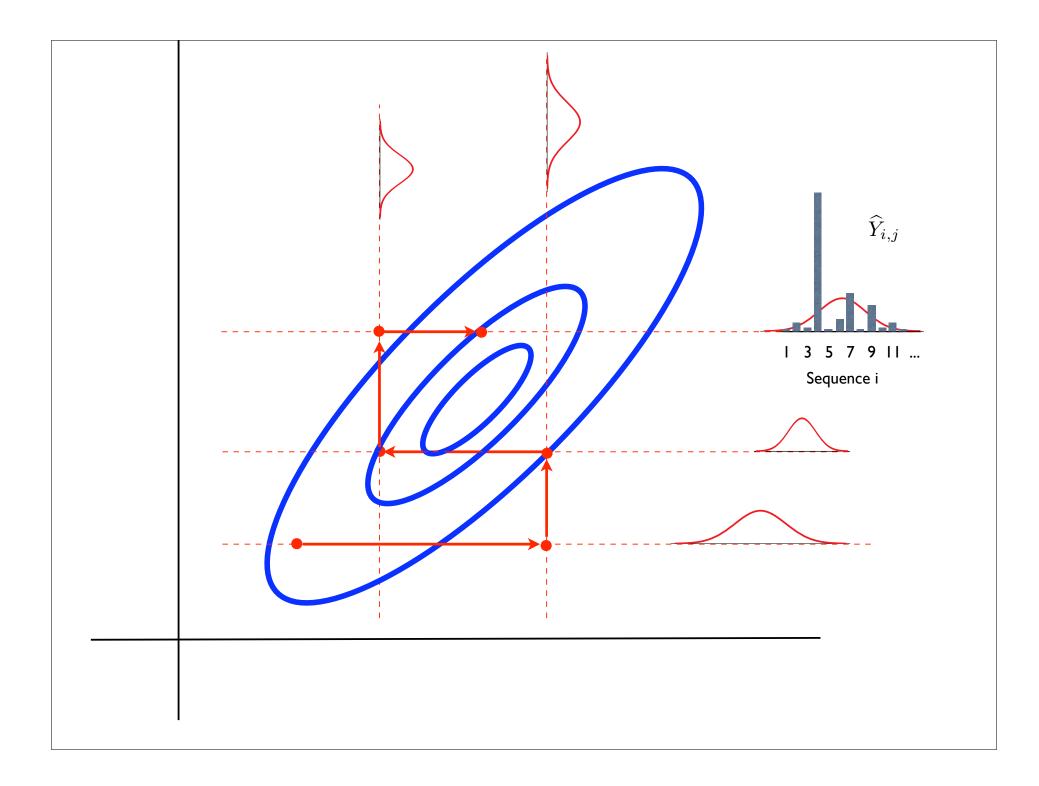
- Approach I: direct integration (rarely solvable analytically, esp. in high dim)
- Approach 2: numerical integration (often difficult, e.g., unstable, esp. in high dim)
- Approach 3: Monte Carlo integration sample  $\vec{x}^{(1)}, \vec{x}^{(2)}, \dots \vec{x}^{(n)} \sim P(\vec{x})$  and average:  $E(f(\vec{x})) \approx \frac{1}{n} \sum_{i=1}^{n} f(\vec{x}^{(i)})$

# Markov Chain Monte Carlo (MCMC)

- Independent sampling also often hard, but not required for expectation
- MCMC  $\vec{X}_{t+1} \sim P(\vec{X}_{t+1} \mid \vec{X}_t)$  w/ stationary dist = P
- Simplest & most common: Gibbs Sampling  $P(x_i \mid x_1, x_2, \dots, x_{i-1}, x_{i+1}, \dots, x_k)$
- Algorithm
  - for  $t = | to \infty$ / \ for i = I to k do :

 $x_{t+1,i} \sim P(x_{t+1,i} \mid x_{t+1,1}, x_{t+1,2}, \dots, x_{t+1,i-1}, x_{t,i+1}, \dots, x_{t,k})$ 

t+1 t



- Input: again assume sequences  $s_1, s_2, ..., s_k$ with one length w motif per sequence
- Motif model: WMM
- Parameters: Where are the motifs? for  $1 \le i \le k$ , have  $1 \le x_i \le |s_i|$ -w+1
- "Full conditional": to calc P(x<sub>i</sub> = j | x<sub>1</sub>, x<sub>2</sub>, ..., x<sub>i-1</sub>, x<sub>i+1</sub>, ..., x<sub>k</sub>) build WMM from motifs in all sequences except *i*, then calc prob that motif in *i<sup>th</sup>* seq occurs at *j* by usual "scanning" alg.

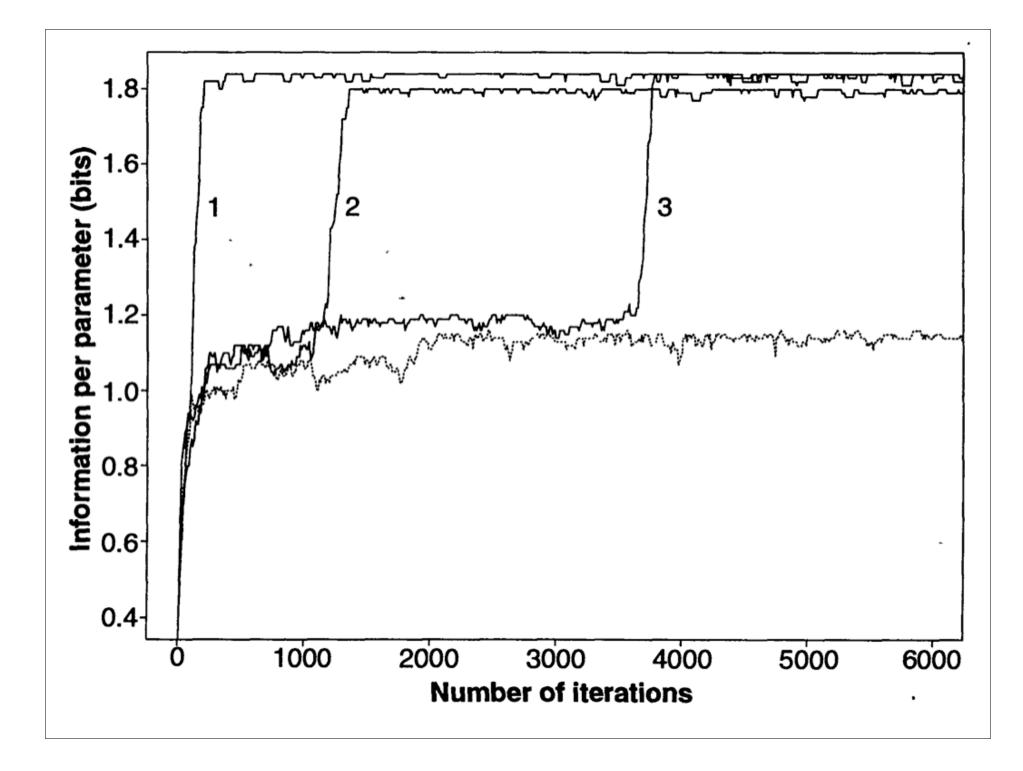
**Overall Gibbs Alg** Randomly initialize  $x_i$ 's for t = | to  $\infty$ for i = 1 to k discard motif instance from s; recalc WMM from rest for  $j = 1 ... |s_i| - w + 1$ Similar to calculate prob that  $i^{th}$  motif is at *j*: MEME, but it bluow  $P(x_i = j \mid x_1, x_2, \dots, x_{i-1}, x_{i+1}, \dots, x_k)$ average over, rather than pick new  $x_i$  according to that distribution sample from

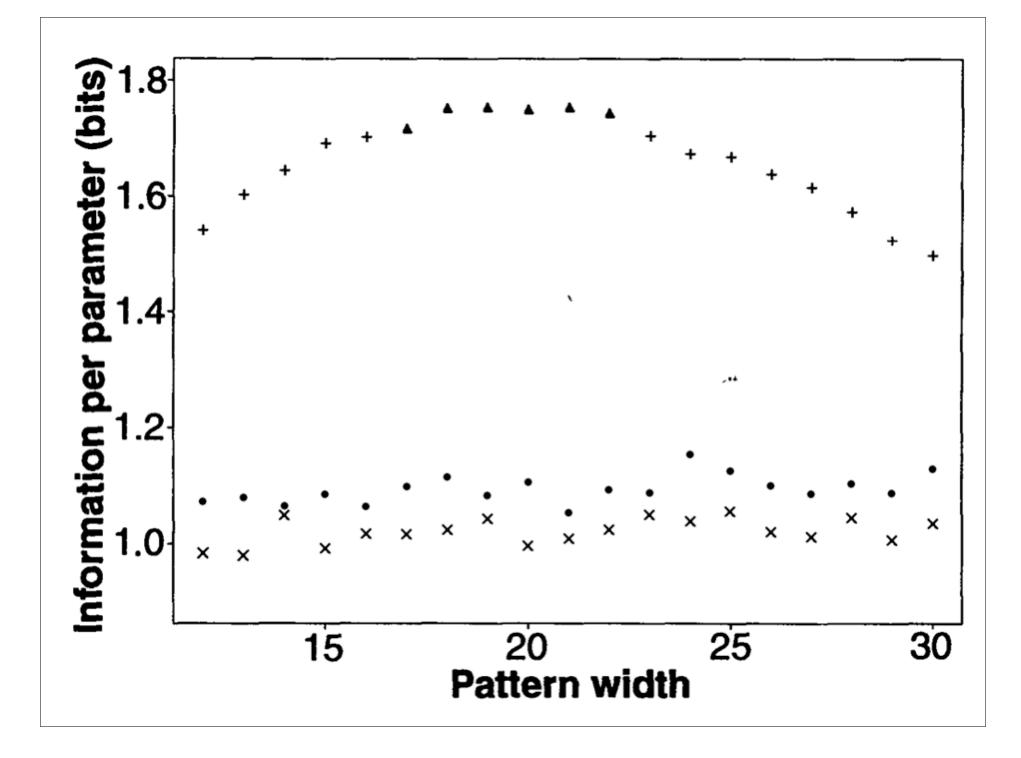
#### Issues

- Burnin how long must we run the chain to reach stationarity?
- Mixing how long a post-burnin sample must we take to get a good sample of the stationary distribution? (Recall that individual samples are not independent, and may not "move" freely through the sample space. Also, many isolated modes.)

#### Variants & Extensions

- "Phase Shift" may settle on suboptimal solution that overlaps part of motif.
   Periodically try moving all motif instances a few spaces left or right.
- Algorithmic adjustment of pattern width: Periodically add/remove flanking positions to maximize (roughly) average relative entropy per position
- Multiple patterns per string



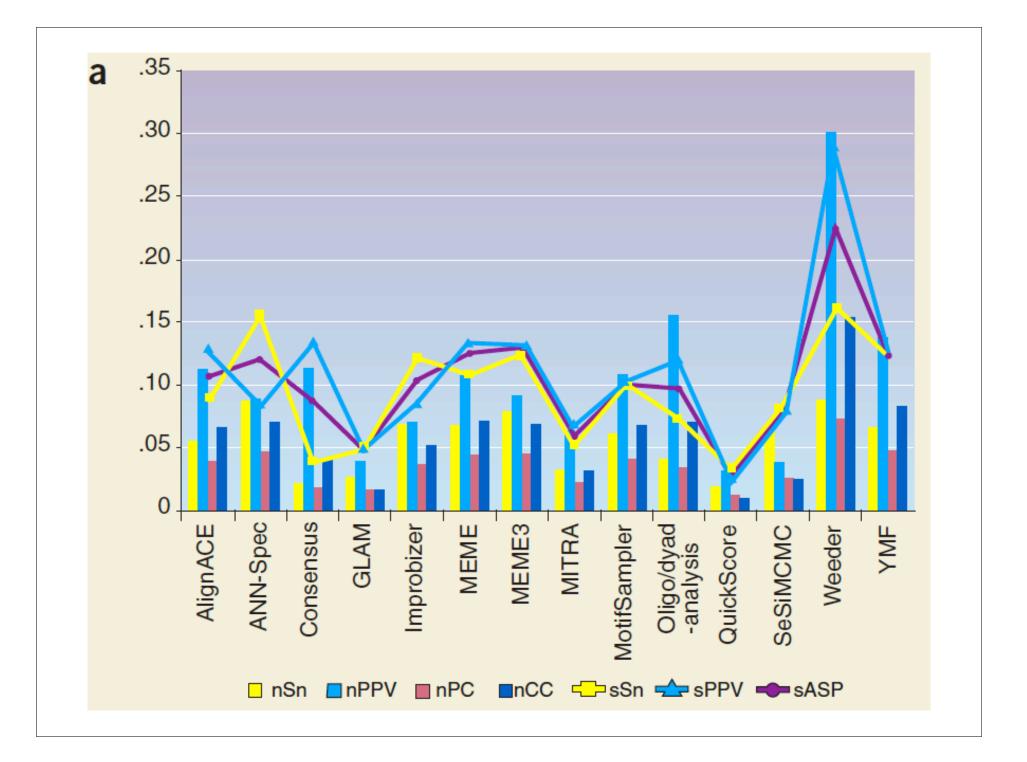


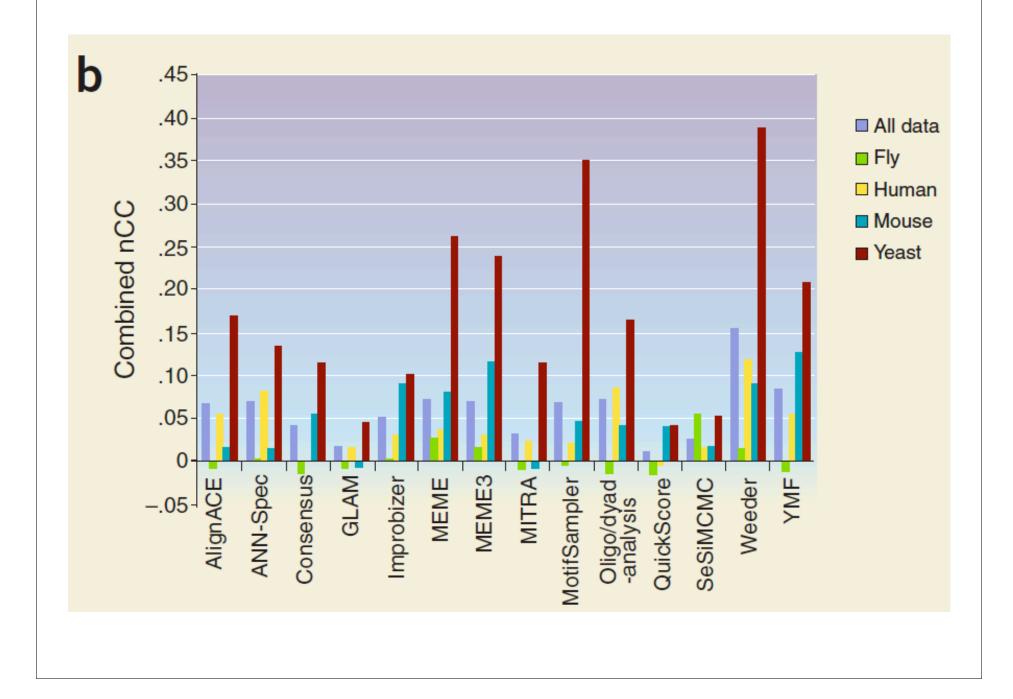
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# Assessing computational tools for the discovery of transcription factor binding sites

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- dozen tools
- Real 'motifs' (Transfac)
- 56 data sets (human, mouse, fly, yeast)
- 'real', 'generic', 'Markov'
- expert users, top prediction only





# Summary

- Important problem: a key to understanding gene regulation
- Hard problem: short, degenerate signals amidst much noise
- Many variants have been tried, for representation, search, and discovery. We looked at only a few:
  - Weight matrix models for representation & search
  - greedy, MEME and Gibbs for discovery
- Still much room for improvement. *Comparative genomics*, i.e. cross-species comparison is very promising